

HFE and hemochromatosis: time to reconsider the diagnostic role of the p.His63Asp variant

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Hemochromatosis, primarily caused by homozygosity for the *HFE* p.Cys282Tyr variant, is the most frequent genetic iron overload disorder in populations of Northern European ancestry. Since the discovery of the *HFE* gene, genetic testing for *HFE*-related hemochromatosis has frequently included both the p.Cys282Tyr and the more common p.His63Asp variants. However, growing evidence indicates that the p.His63Asp variant lacks clear pathogenic significance in most clinical contexts related to iron overload, and its routine reporting may lead to diagnostic confusion and inappropriate management. This article calls for clarification of current genetic testing practices in hemochromatosis, recommending that testing for *HFE*-related hemochromatosis be restricted to the p.Cys282Tyr variant. This position is grounded in current scientific evidence and aims to improve diagnostic accuracy, reduce patient harm, and promote more consistent clinical interpretation.

Introduction

Hemochromatosis is a common inherited metabolic disorder characterized by excessive iron accumulation. Although rare forms of hemochromatosis do exist, *HFE*-related hemochromatosis is now recognized as the most common single-gene disorder in populations of Northern European descent.¹

Given the ongoing uncertainty and debate surrounding the clinical significance of the p.His63Asp variant, a critical clarification of its role in diagnostic and management strategies is warranted. We have conducted a comprehensive review of the current evidence regarding the clinical interpretation of the p.His63Asp variant in hemochromatosis. Based on this analysis, we have formulated a series of focused consensus statements intended to guide its use in diverse clinical settings. These recommendations aim to provide greater clarity for patients, their families, and health care providers. In clinical scenarios that fall outside the scope of these recommendations, referral to a specialist with expertise in iron overload disorders is strongly advised to reduce the risk of misdiagnosis or inappropriate attribution of hemochromatosis.

The history of hemochromatosis spans over a century of evolving concepts, diagnostic criteria, and shifting paradigms in etiology. The condition was first described by A. Trousseau in the 1860s, who observed a triad of diabetes, skin hyperpigmentation, and liver disease, although he did not identify iron

overload as the underlying cause.² In 1889, F. D. von Recklinghausen coined the term “hemochromatosis” and linked the syndrome to iron accumulation, establishing the foundation for viewing it as a systemic iron storage disorder.³ The most comprehensive early description came from J. H. Sheldon in 1935, who published a monograph detailing the clinical and pathologic features and emphasized tissue iron deposition, especially in the liver, as the defining diagnostic criterion.⁴ At that time, diagnosis relied on clinical features (such as bronze diabetes, cirrhosis, and cardiomyopathy) and histopathologic confirmation of iron overload via liver biopsy. It was not until 1975 that Simon et al demonstrated a strong association between idiopathic hemochromatosis and the HLA-A3 and HLA-B7 or B14 alleles, providing the first genetic linkage evidence.^{5,6} Subsequent advances in the 1980s and 1990s shifted diagnostic criteria toward biochemical markers, with elevated serum iron, high transferrin saturation, and increased serum ferritin levels becoming hallmark indicators of the disease.^{7,8} A major breakthrough occurred in 1996 when Feder et al⁹ identified mutations in the “*HFE*” gene as the genetic basis for most adult cases of hemochromatosis, cementing its identity as a Mendelian disorder and establishing genetic testing as a diagnostic cornerstone.

In recent years, the understanding of hemochromatosis has deepened further. It is now recognized as an endocrine liver disease caused by inappropriately low levels of hepcidin, the master regulator of systemic iron homeostasis.¹⁰⁻¹² This reclassification highlights the primary defect in iron sensing and regulation rather than merely iron storage. In clinical practice, invasive liver biopsy, once essential for diagnosis and fibrosis staging, has largely been replaced by noninvasive magnetic resonance imaging (MRI)-based iron quantification and transient elastography, allowing for accurate assessment of hepatic iron burden and fibrosis without procedural risks.¹³ Over time, diagnostic criteria have evolved from symptom-based observation and autopsy findings to biochemical, genetic, and now imaging-based and molecular-functional definitions.¹⁴ This evolution has not been without controversy, as illustrated by the so-called “H63D syndrome,” a concept lacking peer-reviewed clinical evidence, in which overinterpretation of this *HFE* variant led to inflated disease attribution and confusion in both clinical and research settings.

The discovery and recognition of *HFE*

The discovery of mutations in the “*HLA-H*” gene, identified in 1996 through positional cloning,⁹ which was later renamed *HFE* (for “high Fe”¹⁵; OMIM *613609), marked a major breakthrough in understanding the genetic basis of hemochromatosis (OMIM number 235200). In their seminal study, Feder et al⁹ identified 2 variants in the *HLA-H* gene: the p.Cys282Tyr variant (C282Y, rs1800562, NM_000410.4:c.845G>A) and the p.His63Asp variant (H63D, rs1799945, NM_000410.4:c.187C>G). Of 178 well-characterized North American patients with hemochromatosis, 148 were homozygous for the p.Cys282Tyr variant. Among 9 patients heterozygous for p.Cys282Tyr, 8 also carried the p.His63Asp variant, making them compound heterozygotes. One additional patient was homozygous for p.His63Asp.⁹ These 2 variants were found to occur on separate haplotypes, within a genomic linkage region spanning several kilobases, suggesting independent evolutionary origins. Due to its high allele frequency in control populations and similar prevalence (21%) among patients

lacking p.Cys282Tyr, the pathogenic significance of p.His63Asp was uncertain; nevertheless, the observation that 86% of the 178 patients were either homozygous for p.Cys282Tyr or compound heterozygous for p.Cys282Tyr/p.His63Asp was initially interpreted as supporting the involvement of p.His63Asp in the pathogenesis of hemochromatosis. Altogether, this work highlighted a key distinction between disease-associated alleles (individual variants) and disease-associated genotypes (specific combinations) that confer risk. Homozygosity for p.Cys282Tyr mutation emerged as the predominant genotype in hemochromatosis, present in up to 80% to 90% of clinically diagnosed cases in European populations.

An early letter by Beutler,¹⁶ published soon after the discovery of *HFE*, proposed that p.His63Asp could contribute to hemochromatosis in compound heterozygotes, albeit with very low penetrance. Although appropriately cautious, this early framing nonetheless supported the consideration of p.His63Asp as clinically relevant and may have influenced how its significance was subsequently interpreted in diagnostic contexts.

Subsequent population studies of *HFE* variants confirmed that p.Cys282Tyr is the ancestral hemochromatosis mutation. Its high prevalence among Northern Europeans mirrors the geographic distribution of hemochromatosis and supports the hypothesis that the mutation originated in an early Celtic population, later spreading through Celtic migrations facilitated by Viking maritime expertise.¹⁷ In contrast, the p.His63Asp variant shows a broader and less disease-correlated distribution, consistent with it being a more ancient allele. In certain European populations, such as Basques, Catalans, and the Dutch, the p.His63Asp allele frequency exceeds 25%,¹⁸ whereas p.Cys282Tyr occurs at less than half that rate. Notably, in the United Kingdom, p.His63Asp is approximately twice as frequent as p.Cys282Tyr.¹⁹

Statement 1

The original scientific study reporting the strong association between hemochromatosis and the *HFE* genotype containing the p.Cys282Tyr variant in homozygosity also reported additional genotypes containing the most common genetic variant p.His63Asp (Feder et al⁹).

The cellular consequences of the *HFE* variants

The p.Cys282Tyr variant was found to disrupt a key disulfide bond, preventing the formation of the essential tertiary structure of the *HFE* protein and abrogating its association with 2-microglobulin, thereby explaining its absence at the cell surface in individuals homozygous for this variant.^{9,20} The lack of cell surface expression of *HFE* ultimately results in reduced transcription and secretion of hepcidin from hepatocytes. Low circulating levels of hepcidin lead to unchecked release of iron from duodenal enterocytes and reticuloendothelial cells through the transmembrane iron transporter ferroportin.²¹

Contrary to the consequence of the p.Cys282Tyr variant, the functional consequences of the p.His63Asp variant remain obscure. The variant was predicted to lead to an amino acid transition with the putative peptide-binding domain of the protein. This does not lead itself to an immediately plausible mechanism

resulting in the low circulating hepcidin hemochromatosis phenotype. The p.His63Asp variant does not abrogate HFE association with 2-microglobulin, but early studies suggested that it might indirectly alter the affinity of HFE to transferrin receptor.²² Work on mouse models of *HFE*-associated hemochromatosis suggested mild iron overload with p.His67Asp (the mouse equivalent of p.His63Asp in humans) homozygosity and compound heterozygosity with p.Cys294Tyr (the mouse equivalent of p.Cys282Tyr in humans) compared to the mouse equivalent of p.Cys282Tyr homozygosity, but the relevance of these studies to human disease remain unclear.²³

Notably, in humans, p.Cys282Tyr/p.His63Asp compound heterozygotes, who frequently present with additional cofactors such as alcohol use, metabolic syndrome, or steatotic liver disease that can influence iron metabolism, exhibit mildly reduced hepcidin production, although the overall clinical impact remains modest compared with p.Cys282Tyr homozygotes.²⁴

Statement 2

The exact molecular and functional consequences of the p.His63Asp variant remain unknown. Current evidence suggests that the effect, if any, of p.His63Asp-containing genotypes on systemic iron metabolism is minimal.

The limitations of the p.His63Asp variant in contributing to a diagnosis of hemochromatosis

Early analyses of *HFE* mutations in European patients with hemochromatosis revealed an even higher prevalence of p.Cys282Tyr homozygosity than the 83% reported in the original study by Feder et al.⁹ In France,²⁵ United Kingdom,²⁶ Norway,²⁷ Spain,²⁸ and Australia,²⁹ the percentage of well-characterized individual patients who were p.Cys282Tyr homozygous ranged from 87.1% to 100%. Therefore, considering the strong association between p.Cys282Tyr and hemochromatosis, diagnostic laboratories rapidly adopted *HFE* mutation analysis to support early diagnosis of the condition. Despite the weak association of p.His63Asp-containing genotypes with hemochromatosis, *HFE* genotyping, including both the p.Cys282Tyr and p.His63Asp variants, was implemented in routine genetic testing for suspected hemochromatosis.

In a subsequent report,³⁰ the relative risk for hemochromatosis in individuals with the p.Cys282Tyr/p.His63Asp compound heterozygous genotype was only 0.5%, compared with the more highly penetrant p.C282Y homozygous genotype. Therefore, it was concluded that despite analyses indicating a potential association of certain p.His63Asp-containing genotypes and disease, there was a need to more precisely define the absolute risk linked to each *HFE* genotype, considering factors such as age, sex, and environmental factors.³⁰ Numerous subsequent case-control studies reported that only a relatively small proportion of patients with hemochromatosis carried genotypes involving the p.His63Asp variant. A large meta-analysis including 202 studies with 66 263 cases and 226 515 controls found no statistically significant association between genotypes containing the p.His63Asp variant and liver disease or any other clinical outcomes typically associated with hemochromatosis, including heart disease, arthropathy, and diabetes.³¹

True iron overload associated with hemochromatosis requires documentation of elevation of the transferrin saturation combined with an elevation of serum ferritin levels. Although elevated transferrin saturation is widely considered the most sensitive marker of the hemochromatosis phenotype, it is subjected to significant biological variability, influenced by variables such as circadian timing and fasting state at the time of sample collection. Serum ferritin is a nonspecific marker, because studies indicate that <10% of elevated ferritin levels are attributed to hemochromatosis, with most cases instead associated with inflammation, chronic alcohol consumption, or hepatic steatosis.³²

According to European Association for the Study of the Liver guidelines,¹⁴ p.His63Asp genotyping is not routinely recommended but may be considered in specific clinical contexts, because its clinical relevance remains controversial.

An individual with a low-risk hemochromatosis genotype (including p.Cys282Tyr/p.His63Asp compound heterozygosity and p.His63Asp homozygosity) and a raised ferritin should not be considered to have hemochromatosis without further evidence of increased tissue iron overload, ideally confirmed prospectively by positive MRI or, in the absence of MRI, by measurements confirming elevation of body iron stores. The potential phenotypic expression in compound heterozygotes may be influenced by additional factors, including alcohol use, metabolic syndrome, and steatotic liver disease, which can modify the severity and onset of iron-related abnormalities.^{33,34}

Statement 3

The p.His63Asp variant when present in compound heterozygosity with p.Cys282Tyr can be considered to confer a risk for mild iron overload, commonly in association with other risk factors (including alcohol use, metabolic syndrome, and steatotic liver disease). It is not considered to be a disease-causing genotype on its own.

Testing for p.His63Asp has limited utility in evaluation of patients with significant iron overload

Since the identification of the *HFE* gene, it has been widely recognized that homozygosity for p.Cys282Tyr mutation is the most common risk factor for iron overload in humans. Rarer forms of genetic iron overload are recognized, and their phenotype may vary significantly from *HFE*-associated hemochromatosis. Such patients require specialist assessment and management. This should include quantification of liver iron concentration by MRI and assessment of cardiac and endocrine functions.^{13,35} Genetic analysis in such instances should include not only testing for p.His63Asp but should include sequencing the entire *HFE* gene and other genes involved in iron metabolism (*HJV*, *TFR2*, *HAMP*, and *SLC40A1*).³⁶ Clearly, this is beyond the capabilities of most first-line routine clinical testing services, highlighting the necessity for specialist consultation in a more in-depth second-line investigation.

Statement 4

Regardless of their p.His63Asp status, patients with “unexplained” significant tissue iron overload, demonstrated either by a direct measure of liver iron concentration (prospective determination) or by quantitative phlebotomy (retrospective determination), and

without *HFE* p.Cys282Tyr homozygosity should be tested for other pathogenic variants in the *HFE* and non-*HFE* genes.

Assessment of p.His63Asp has no role in screening and case finding

Cascade screening has been recommended in international guidelines and a practiced feature of hemochromatosis care since its hereditary nature was established. The identification of the *HFE* gene, particularly the p.Cys282Tyr variant, greatly expanded the potential for early genetic testing in siblings and offspring of individuals with p.Cys282Tyr homozygosity. Genetic-based testing in families in which the affected proband has a genotype different from p.Cys282Tyr homozygosity (including p.Cys282Tyr/p.His63Asp compound heterozygosity and p.His63Asp homozygosity) can be considered “uninformative” (ie, the predictive value of testing is too low to be of value). Therefore, assessment of p.His63Asp for family cascade or population screening is not commonly practiced and not recommended.³⁷

The largest population study of the *HFE* variants, to our knowledge, analyzed p.Cys282Tyr and p.His63Asp in >450 000 UK residents followed up for 7 years.¹⁹ Although p.Cys282Tyr homozygosity was strongly associated with hemochromatosis, liver disease, liver cancer, and joint and neurological conditions, the risk in p.Cys282Tyr/p.His63Asp compound heterozygotes was modest, and no increased morbidity was observed in p.His63Asp homozygotes.^{19,38} After 13 years, the cumulative hemochromatosis diagnosis in nearly 5000 male compound heterozygotes was only 5.4% by the age of 80, compared to 56.4% in p.Cys282Tyr homozygotes.³⁹ Similar studies in Australia^{40,41} and Denmark^{42,43} confirmed no increased risk of liver disease, diabetes, or iron overload in individuals with p.His63Asp-containing genotypes.

In countries where hemochromatosis and the p.Cys282Tyr variant have a high prevalence, there has been a longstanding debate regarding the potential value of population screening.⁴⁴ To date, no country has yet adopted population screening, and although there is an increasing recognition that testing for p.Cys282Tyr might be a model to consider, there is no evidence that testing for p.His63Asp would contribute to such a program. There is no evidence to suggest that prospective testing for p.His63Asp in individuals without evidence of iron overload is beneficial.

Statement 5

The inclusion of the p.His63Asp variant for assessing hemochromatosis risk in asymptomatic individuals (including cascade or family testing and population screenings) offers no clinical benefit and is not recommended.

Research testing and direct-to-consumer genetic tests

Given the prevalence of hemochromatosis and of genotypes bearing the p.Cys282Tyr variant in certain populations, there is an increasing demand for genetic variant analysis from clinicians in both secondary and primary care settings. Genetic risk must be communicated to affected individuals. Any such communication needs to be clear, evidence based, and clinically balanced. Moreover, a growing number of individuals are subjected to genetic testing outside the framework and oversight of their

established health care providers. Many people currently receive highly technical and personalized genetic data directly from research bodies and commercial organizations. In many countries, this practice is being promoted by industry leaders, offering to cover more and more genes as well as partnering with patient associations to expand access. Results inferring a disease risk can lead to anxiety and distress for patients, often resulting in additional consultations and medical tests.

The American College of Medical Genetics and Genomics define only p.Cys282Tyr homozygosity as an “actionable” incidental genetic finding, that is, a finding of sufficient significance to warrant communication to the patient/participant/customer along with a clinical assessment and management.⁴⁵ Given the high prevalence of p.His63Asp across the world, the unlimited testing of this variant creates a huge potential for misdiagnosis and even unnecessary treatment. This should be consistently resisted,⁴⁶ and any laboratory performing and reporting on genetic information (including research and commercial partners) must acknowledge their clinical responsibility.

Conclusion and recommendations

The p.His63Asp variant in *HFE* remains a genetic variant of interest. However, there is insufficient evidence to justify the continued routine reporting of this variant in all referrals. This variant has never been informative in the context of family analysis, and therefore, it should not be reported in cascade screening. Testing for p.His63Asp may be considered as part of second-line genetic analysis in patients with unequivocal evidence of iron overload,³⁷ alongside analysis of other *HFE* regions and non-*HFE* hemochromatosis genes (*HJV*, *TFR2*, *HAMP*, and *SLC40A1*), preferably in specialized centers with expertise in interpreting pathogenic variants in hemochromatosis.

Without more selective and clinically informed reporting, colleagues will continue to be overwhelmed with patients presenting modest elevations of serum ferritin and what are essentially low- or no-risk genotypes (ie, as p.Cys282Tyr/p.His63Asp compound heterozygotes or p.His63Asp homozygotes). We have no evidence that these genotypes alone confer serious risk or that the individual would ever benefit from iron reduction/venesection treatment.^{19,39}

We have a responsibility to make an early yet accurate diagnosis of hemochromatosis before recommending treatment. A false diagnosis, such as may occur in any individual with isolated hyperferritinemia and a low-risk *HFE* genotype (but without compelling evidence of iron overload), is unacceptable. It may lead to unnecessary venesection, as well as potential stigma and psychological harm. Given the wide range of nonspecific symptoms associated with hemochromatosis and iron overload, patients may mistakenly attribute new symptoms to their (incorrect) diagnosis. This misattribution can lead to delays in reporting symptoms, undergoing proper investigation and receiving timely diagnosis of other potentially serious health conditions.

In the context of emerging targeted molecular therapies for hemochromatosis and to rigorously assess the long-term efficacy of such therapies in broader populations, it is essential to focus on patient group at greatest risk of serious disease. An early

“proof-of-concept” study evaluating the hepcidin mimetic rusfertide (PTG-300, Protagonist Inc) showed promising tolerance and potential benefit.⁴⁷ However, of the 16 participants, only 5 were confirmed p.Cys282Tyr homozygotes, whereas others carried other genotypes containing the p.His63Asp variant. Future trials should therefore concentrate on individuals homozygous for p.Cys282Tyr, who represent the group most likely to benefit from targeted intervention.

Overall, in patients with confirmed tissue iron overload, testing for the HFE p.His63Asp variant may have limited clinical utility, especially when broader genetic and environmental factors are considered. Routine reporting of this variant, particularly in individuals without clear evidence of iron overload, offers no diagnostic benefit and should be discouraged to prevent misinterpretation and unnecessary clinical actions.

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References

1. Barton JC, Parker CJ. HFE-related hemochromatosis. In: Adam MP, Mirzaa GM, Pagon RA, et al, eds. *GeneReviews*. University of Washington; 2008. Updated 11 April 2024. Accessed 1 December 2025. <https://www.ncbi.nlm.nih.gov/books/NBK1440/>
2. Trosseau A. In: *Glycosurie; Diabete Sucré*. 2nd ed. Clinique Méd de l'Hotel-Dieu de Paris; 1865.
3. von Recklinghausen DF. Über Hämochromatose. *Tagblatt der Versammlung Deutscher Naturforscher und Ärzte in Heidelberg*. 1889;62:324-325.
4. Sheldon JH. *Haemochromatosis*. 1st ed. Oxford University Press; 1935.
5. Simon M, Bourel M, Fauchet R, Genetet B. Association of HLA-A3 and HLA-B14 antigens with idiopathic haemochromatosis. *Gut*. 1976;17(5):332-334.
6. Simon M, Bourel M, Genetet B, Fauchet R. Idiopathic hemochromatosis. Demonstration of recessive transmission and early detection by family HLA typing. *N Engl J Med*. 1977;297(19):1017-1021.
7. Bothwell TH, Charlton RW. Historical overview of hemochromatosis. *Ann N Y Acad Sci*. 1988;526:1-10.
8. Bothwell TH, Jacobs P, Torrance JD. Studies on the behaviour of transferrin in idiopathic haemochromatosis. *S Afr J Med Sci*. 1962;27:35-39.
9. Feder JN, Gnirke A, Thomas W, et al. A novel MHC class I-like gene is mutated in patients with hereditary haemochromatosis. *Nat Genet*. 1996;13(4):399-408.
10. Bridle KR, Frazer DM, Wilkins SJ, et al. Disrupted hepcidin regulation in HFE-associated haemochromatosis and the liver as a regulator of body iron homeostasis. *Lancet*. 2003;361(9358):669-673.
11. Pietrangelo A. Hemochromatosis: an endocrine liver disease. *Hepatology*. 2007;46(4):1291-1301.
12. Nemeth E, Ganz T. Hepcidin and iron in health and disease. *Annu Rev Med*. 2023;74:261-277.
13. Henninger B, Alustiza J, Garbowski M, Gandon Y. Practical guide to quantification of hepatic iron with MRI. *Eur Radiol*. 2020;30(1):383-393.
14. European Association for the Study of the Liver. EASL clinical practice guidelines on haemochromatosis. *J Hepatol*. 2022;77(2):479-502.
15. Bodmer JG, Parham P, Albert ED, Marsh SG. Putting a hold on "HLA-H". The WHO Nomenclature Committee for factors of the HLA system. *Nat Genet*. 1997;15(3):234-235.
16. Beutler E. The significance of the 187G (H63D) mutation in hemochromatosis. *Am J Hum Genet*. 1997 Sep;61(3):762-764.
17. Distante S, Robson KJ, Graham-Campbell J, Arnaiz-Villena A, Brissot P, Worwood M. The origin and spread of the HFE-C282Y hemochromatosis mutation. *Hum Genet*. 2004 Sep;115(4):269-279.

18. Merryweather-Clarke AT, Pointon JJ, Shearman JD, Robson KJ. Global prevalence of putative haemochromatosis mutations. *J Med Genet.* 1997;34(4):275-278.
19. Pilling LC, Tamosauskaite J, Jones G, et al. Common conditions associated with hereditary haemochromatosis genetic variants: cohort study in UK Biobank. *BMJ.* 2019;364:k5222.
20. Feder JN, Tsuchihashi Z, Irrinki A, et al. The hemochromatosis founder mutation in HLA-H disrupts beta2-microglobulin interaction and cell surface expression. *J Biol Chem.* 1997;272(22):14025-14028.
21. Muckenthaler MU. How mutant HFE causes hereditary hemochromatosis. *Blood.* 2014;124(8):1212-1213.
22. Feder JN, Penny DM, Irrinki A, et al. The hemochromatosis gene product complexes with the transferrin receptor and lowers its affinity for ligand binding. *Proc Natl Acad Sci U S A.* 1998;95(4):1472-1477.
23. Tomatsu S, Orii KO, Fleming RE, et al. Contribution of the H63D mutation in HFE to murine hereditary hemochromatosis. *Proc Natl Acad Sci U S A.* 2003;100(26):15788-15793.
24. Piperno A, Girelli D, Nemeth E, et al. Blunted hepcidin response to oral iron challenge in HFE-related hemochromatosis. *Blood.* 2007 Dec 1;110(12):4096-4100.
25. Jouanolle AM, Gandon G, Jézéquel P, et al. Haemochromatosis and HLA-H. *Nat Genet.* 1996;14(3):251-252.
26. The UK Haemochromatosis Consortium. A simple genetic test identifies 90% of UK patients with haemochromatosis. *Gut.* 1997;41(6):841-844.
27. Bell H, Berg JP, Undlien DE, et al. The clinical expression of hemochromatosis in Oslo, Norway. Excessive oral iron intake may lead to secondary hemochromatosis even in HFE C282Y mutation negative subjects. *Scand J Gastroenterol.* 2000;35(12):1301-1307.
28. Sánchez M, Bruguera M, Bosch J, Rodés J, Ballesta F, Oliva R. Prevalence of the Cys282Tyr and His63Asp HFE gene mutations in Spanish patients with hereditary hemochromatosis and in controls. *J Hepatol.* 1998;29(5):725-728.
29. Jazwinska EC, Cullen LM, Busfield F, et al. Haemochromatosis and HLA-H. *Nat Genet.* 1996;14(3):249-251.
30. Risch N. Haemochromatosis, HFE and genetic complexity. *Nat Genet.* 1997;17(4):375-376.
31. Ellervik C, Birgens H, Tybjaerg-Hansen A, Nordestgaard BG. Hemochromatosis genotypes and risk of 31 disease endpoints: meta-analyses including 66,000 cases and 226,000 controls. *Hepatology.* 2007;46(4):1071-1080.
32. Hearnshaw S, Thompson NP, McGill A. The epidemiology of hyperferritinaemia. *World J Gastroenterol.* 2006;12(36):5866-5869.
33. Allen KJ, Gurrin LC, Constantine CC, et al. Iron-overload-related disease in HFE hereditary hemochromatosis. *N Engl J Med.* 2008;358(3):221-230.
34. Wallace DF, Subramaniam VN. Co-factors in liver disease: the role of HFE-related hereditary hemochromatosis and iron. *Biochim Biophys Acta.* 2009 Jul;1790(7):663-670.
35. Reeder SB, Yokoo T, França M, et al. Quantification of liver iron overload with MRI: review and guidelines from the ESGAR and SAR. *Radiology.* 2023;307(1):e221856.
36. Girelli D, Busti F, Brissot P, Cabantchik I, Muckenthaler MU, Porto G. Hemochromatosis classification: update and recommendations by the BIOIRON Society. *Blood.* 2022;139(20):3018-3029.
37. Porto G, Brissot P, Swinkels DW, et al. EMQN best practice guidelines for the molecular genetic diagnosis of hereditary hemochromatosis (HH). *Eur J Hum Genet.* 2016;24(4):479-495.
38. Atkins JL, Pilling LC, Heales CJ, et al. Hemochromatosis mutations, brain iron imaging, and dementia in the UK Biobank cohort. *J Alzheimers Dis.* 2021;79(3):1203-1211.
39. Lucas MR, Atkins JL, Pilling LC, Shearman JD, Melzer D. HFE genotypes, haemochromatosis diagnosis and clinical outcomes at age 80 years: a prospective cohort study in the UK Biobank. *BMJ Open.* 2024;14(3):e081926.
40. Gurrin LC, Bertalli NA, Dalton GW, et al; HealthIron Study Investigators. HFE C282Y/H63D compound heterozygotes are at low risk of hemochromatosis-related morbidity. *Hepatology.* 2009;50(1):94-101.
41. Zaloumis SG, Allen KJ, Bertalli NA, et al; HealthIron Study Investigators. Natural history of HFE simple heterozygosity for C282Y and H63D: a prospective 12-year study. *J Gastroenterol Hepatol.* 2015;30(4):719-725.
42. Mottelson M, Glenthøj A, Nordestgaard BG, et al. Iron, hemochromatosis genotypes, and risk of infections: a cohort study of 142 188 general population individuals. *Blood.* 2024;144(7):693-707.
43. Mottelson M, Helby J, Nordestgaard BG, et al. Mortality and risk of diabetes, liver disease, and heart disease in individuals with haemochromatosis HFE C282Y homozygosity and normal concentrations of iron, transferrin saturation, or ferritin: prospective cohort study. *BMJ.* 2024;387:e079147.
44. Delatycki MB, Allen KJ. Population screening for hereditary haemochromatosis—should it be carried out, and if so, how? *Genes.* 2024;15(8):967.
45. Miller DT, Lee K, Abul-Husn NS, et al; ACMG Secondary Findings Working Group. Electronic address: documents@acmg.net. ACMG SF v3.2 list for reporting of secondary findings in clinical exome and genome sequencing: a policy statement of the American College of Medical Genetics and Genomics (ACMG). *Genet Med.* 2023;25(8):100866.
46. Mahase E, Iacobucci G. Genetic test “screening campaign” may be causing unnecessary alarm, experts warn. *BMJ.* 2023;381:1264.
47. Kowdley KV, Modi NB, Peltekian K, et al. Rusfertide for the treatment of iron overload in HFE-related haemochromatosis: an open-label, multicentre, proof-of-concept phase 2 trial. *Lancet Gastroenterol Hepatol.* 2023;8(12):1118-1128.